# A LIGNAN GLUCOSIDE FROM EUPHRASIA ROSTKOVIANA\*

# OSAMA SALAMA,† RATAN K. CHAUDHURI and OTTO STICHER

Eidgenössische Technische Hochschule, Pharmazeutisches Institut, ETH-Zentrum, 8092 Zürich, Switzerland

(Received 12 March 1981)

**Key Word Index**—*Euphrasia rostkoviana*; Scrophulariaceae; structure elucidation; lignan glucoside; dihydrobenzofuran lignan; dehydrodiconiferyl alcohol-4-β-D-glucoside; <sup>13</sup>C NMR.

**Abstract**—A new lignan glucoside has been isolated from *Euphrasia rostkoviana*. Its structure was elucidated by spectroscopic means and by correlation with dehydrodiconiferyl alcohol to be dehydrodiconiferyl alcohol-4- $\beta$ -D-glucoside.

## INTRODUCTION

During the isolation of iridoids from Euphrasia rostkoviana [2], we encountered a non-iridoid glycoside that has now been identified as a new lignan glucoside. In this paper we describe the isolation and structure determination of this compound, namely dehydrodiconiferyl alcohol-4- $\beta$ -D-glucoside.

## RESULTS AND DISCUSSION

The water-soluble part of *E. rostkoviana* was filtered through a column of neutral alumina and the water eluates were lyophilized and fractionated over a Si gel column. The appropriate eluates (TLC) were combined and further chromatography over  $C_{18}$  reversed phase column gave pure dehydrodiconiferyl alcohol-4- $\beta$ -D-glucoside (1).

The molecular formula  $C_{26}H_{32}O_{11}$  of 1 is based on field desorption—mass spectrometry (M<sup>+</sup>, 520), the number of signals in the PND <sup>13</sup>C NMR and their multiplicities in the SFORD and integration of the <sup>1</sup>H NMR spectra. The <sup>1</sup>H NMR (360 MHz) spectrum showed five aromatic protons, the coupling constants of which gave the substitution pattern indicated in the formula, two olefinic protons (H- $\alpha$ ' and H- $\beta$ ') which appeared as an AB part of an ABX<sub>2</sub> system, two OMe groups and an anomeric  $\beta$ -D-glucose proton. The large coupling constant ( $J_{1'',2''}=7.5$  Hz) proves the  $\beta$ -configuration of the anomeric centre. The region of the remaining glucoside protons had the expected splitting

1  $R = \beta$ -D-Glucose; R' = H

2 R = Tetraacetyl- $\beta$ -D-glucose; R' = Ac

3 R = R' = H

patterns and integral value. The <sup>1</sup>H NMR spectrum of the hexacetate 2 was similarly related. From the above data, structure 1 (disregarding the site of glucosidation) can be proposed for the new compound.

Further confirmation of the structure of 1 as well as the site of glucosidation came from an analysis of its <sup>13</sup>C NMR spectrum which showed, apart from the six signals due to  $\beta$ -D-glucose, signals corresponding to 18 carbon atoms (excluding the signals due to  $2 \times OCH_3$ ), consistent with a lignan structure. The assignment of the signals based on the literature precedent [3] are given in Table 1. The  $\delta$  values quoted for 3 from ref. [3] are for a solution in acetone-d<sub>6</sub>/D<sub>2</sub>O and are comparable with spectra recorded in CD<sub>3</sub>OD. Out of 18 signals in 3, 15 signals are virtually identical to signals observed in the <sup>13</sup>C NMR of 1. An expected upfield shift of the signal of C-4 in 1, due to the glucosidation, is not observed in respect to 3. However, significant downfield shifts of the signals of the ortho- (C-3 and C-5) and para- (C-1) related carbons are observed, thereby locating [4, 5] the glucose moiety at C-4. Consequently, the compound 1 is dehydrodiconiferyl alcohol-4- $\beta$ -D-glucoside.

Table 1. <sup>13</sup>C NMR data of dehydrodiconiferyl alcohol-4-β-D-glucoside (1) and its aglycone 3

C atom	1*	3†	C atom	1*	3†
1	137.5	134.3	1'	129.7	130.4
2	110.9	110.7	2'	111.7	111.8
3	150.4	148.6	3′	145.0	145.1
3-OMe	56.4	+	3'-OMe	56.4	<b>+</b>
4	147.1	147.3	4′	148.7	148.9
5	117.5	115.8	5′	132.2	132.0
6	119.5	119.5	6′	116.1	116.2
α	88.4	88.3	α'	131.6	130.9
β	54.7	54.6	β'	127.3	128.0
γ	64.5	64.6	γ'	63.5	63.3
i''	102.2		4''	70.8	_
2′′	74.4		5''	77.3	_
3′′	77.6		6′′	62.1	_

<sup>\*</sup>Run at 25.2 MHz in CD<sub>3</sub>OD with TMS as an internal standard.

<sup>\*</sup>Part 2 in the series "Glycosides of *Euphrasia* species". For Part 1 see ref. [1].

<sup>†</sup>Part of a Ph.D. thesis to be submitted to the ETH, Zürich.

<sup>†</sup>Data from ref. [3], in  $Me_2CO-d_6-D_2O$  (9:1).

<sup>‡</sup>Not mentioned in lit. [3].

2604

Compound 1 has not been reported before in nature although its aglycone 3 was known long ago [6]. Also, this is the first demonstration of the occurrence of a lignan in the genus *Euphrasia*.

## EXPERIMENTAL

The general methods are the same as reported in ref. [7]. Extraction. Dried and milled whole plant (1kg) of E. rostkoviana Hayne, available commercially from Siegfried AG (Lot No. 19279), Zofingen, Switzerland, was extracted with MeOH at 40° (4 × 51.). After concn of the combined extracts in vacuo, H<sub>2</sub>O (1.51.) was added and the H<sub>2</sub>O-insoluble material removed by filtration through Celite. The filtrate was extracted with petrol (60 80°, 4 × 11.) and the petrol-soluble part rejected. The aq. layer was concd (200 ml) and the aq. conct filtered through a prewashed (H<sub>2</sub>O) neutral Al<sub>2</sub>O<sub>3</sub> (500 g) column eluting with H<sub>2</sub>O. The aq. eluate was concd and lyophilized to give the crude glycoside fraction (45 g). A portion of the residue (25 g) was chromatographed over Si gel (400 g) eluting with CH<sub>2</sub>Cl<sub>2</sub>. MeOH-H<sub>2</sub>O, 40:10:1 (31.), 70:30:3 (31.), 14:10:1 (21.) and five fractions A-E were collected.

Isolation of dehydrodiconiferyl alcohol-4-β-D-glucoside (1). Fraction B (1.8 g) was further chromatographed over Si gel (100 g) eluting with EtOAc-n-PrOH- $H_2O$  (4:2:7, upper layer) and two fractions  $B_1$  and  $B_2$  were collected. Low pressure liquid chromatography (reversed phase  $C_{18}$  with MeOH- $H_2O$ , 1:3) of  $B_1$  (200 mg) gave pure 1 (ca. 0.013%), [α] $_0^{20}$  - 71.2% (c = 0.56, MeOH). UV  $\lambda_{\max}^{\text{MeOH}}$  nm: 274 (log  $\varepsilon$  = 4.04) and 217 (log  $\varepsilon$  = 4.35). IR  $\nu_{\max}$  cm $^{-1}$ : 3380 (br OH), 1600, 1500, 1460 (aromatic ring). HNMR (360 MHz, DMSO- $d_0$ ): δ 7.07 (1 H, d, J = 8.5 Hz, H-5), 6.96 (1 H, d, J =  $\sim$  2 Hz, H-2'), 6.93 (2 H, J =  $\sim$  2 Hz, H-2/6'), 6.84 (1 H, dd, J = 8.5/2 Hz, H-6), 6.47 (1 H, d, J = 16 Hz, H- $\alpha$ '), 6.23 (1 H, dt, J = 16/5 Hz, H- $\beta$ '), 5.51 (1 H, d, J = 6.5 Hz, H- $\alpha$ ) 4.88 (1 H, d, J = 7.5 Hz, H-1"), 3.82 (3 H, s, 3-OMe), 3.75 (3 H, s, 3'-OMe),  $\sim$  3.75 (1 H, partly merged with the

3'-OMe signal, H- $\beta$ ), 3.65 (2 H, m, 2H- $\gamma$ '), 3.44 (2J, dd, J = 6/6 Hz, 2H- $\gamma$ ); <sup>13</sup>C NMR (Table 1). EIMS 70 eV m/z (rel. int.): 520 [M] <sup>+</sup> (1.6), 358 [M - 163] <sup>+</sup> with H transfer (17), 342 [M - 179] <sup>+</sup> (28), 340 [M - (163 + H<sub>2</sub>O)] <sup>+</sup> (28), 324 [M - (179 + H<sub>2</sub>O)] <sup>+</sup> (56), 312 [M <sup>+</sup> - (179 + 30)] <sup>+</sup> (28), 151 (11), 137 (28), 115 (17), 85 (22), 73 (89), 60 (100), 43 (83), 30 (33), 29 (89), 18 (39).

Dehydrodiconiferyl alcohol-4-β-D-glucoside-hexaacetate (2). Acetylation of 1 with Ac<sub>2</sub>O-pyridine at room temp for 18 hr followed by usual work-up gave the hexaacetate 2 as an amorphous powder,  $[\alpha]_{C}^{20} - 2^{\circ}$  (c = 0.87, CHCl<sub>3</sub>); <sup>1</sup>H NMR (CDCl<sub>3</sub>, 100 MHz):  $\delta$  7.06 (1 H, d, J = 8 Hz, H-5), 6.96–6.82 (4 H, H-2, 2′, 5,6), 6.58 (1 H, d, J = 16 Hz, H-α′), 6.12 (1 H, dt, J = 16/6 Hz, H-β′), 5.48 (1 H, d, J = 7 Hz, H-α), 4.69 (2 H, d, J = 6 Hz, H-γ′), 4.4–4.14 (m, 2 H-γ′), 3.90 (3 H, s, 3-OMe), 3.77 (3 H, s, 3'-OMe), ~3.74 (1 H, m, partly merged with the 3'-OMe signal, H-β), 2.12–1.96 (18 H, 6 × OAc).

Acknowledgements—This work was supported by research grants of the Swiss Federal Institute of Technology (ETH) and the Swiss National Science Foundation. Thanks are due to Dr. T. Winkler, Ciba-Geigy, Basel, for some helpful discussions.

## REFERENCES

- 1. Sticher, O. and Salama, O. (1981) Helv. Chim. Acta 64, 78.
- 2. Sticher, O. and Salama, O. (1980) Planta Med. 39, 269.
- Lüdemann, H.-D. and Nimz, H. (1974) Makromolek. Chem. 175, 2393.
- 4. Cussans, N. J. and Huckerby, T. N. (1975) Tetrahedron 31,
- Markham, K. R., Ternai, B., Stanley, R., Geiger, H. and Mabry, T. (1978) Tetrahedron 34, 1389.
- Weinges, K., Müller, R., Kloss, P. and Jaggy, H. (1970) Ann. Chem. 736, 170.
- Chaudhuri, R. K. and Sticher, O. (1980) Helv. Chim. Acta 63, 117.